

PCR Primers Design

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Testing Primer Specificity

- ▶ Most important factor affecting PCR: Choosing **appropriate primers**
- ▶ To amplify exactly the intended target and avoiding undesired amplification
- ▶ Primers should not have **matches** to other targets in certain orientations and within certain distances

Process of designing specific primers:

- ▶ Generating primers flanking the target region (manually or using software tools)
- ▶ Searching primers against an appropriate nucleotide sequence database
 - to examine the potential targets
 - to examine many details between primers and targets, such as:
 - number and the positions of matched bases
 - primer orientations and distance between forward and reverse primers
- ▶ **Target specificity**: critical primer property

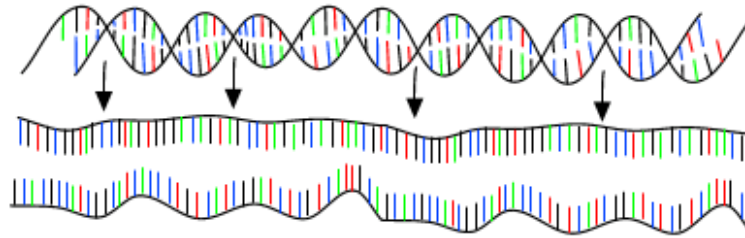
Primers dictate the successfulness of a PCR

PCR : Polymerase Chain Reaction

30 - 40 cycles of 3 steps :

Step 1 : denaturation

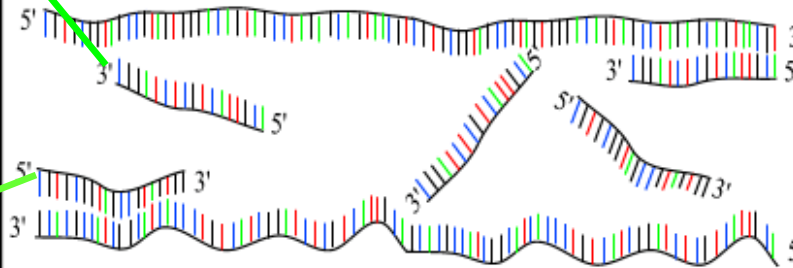
1 minut 94 °C



Step 2 : annealing

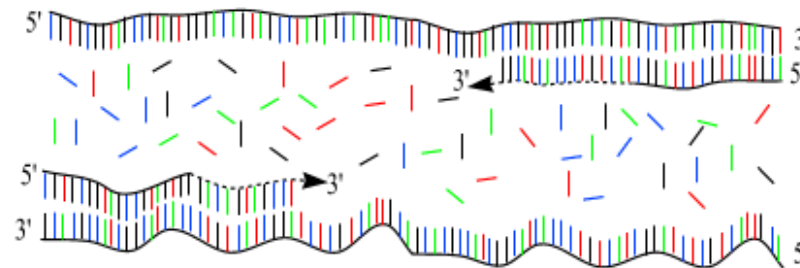
45 seconds 54 °C

forward and reverse primers !!!



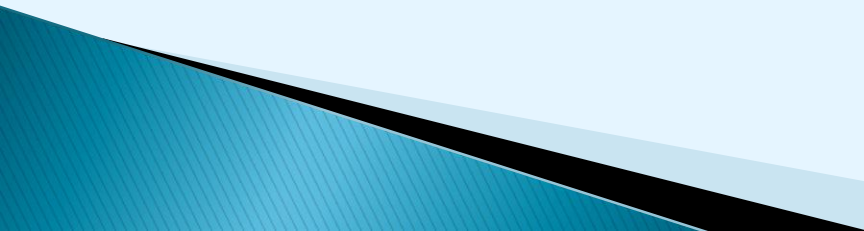
Step 3 : extension

2 minutes 72 °C
only dNTP's

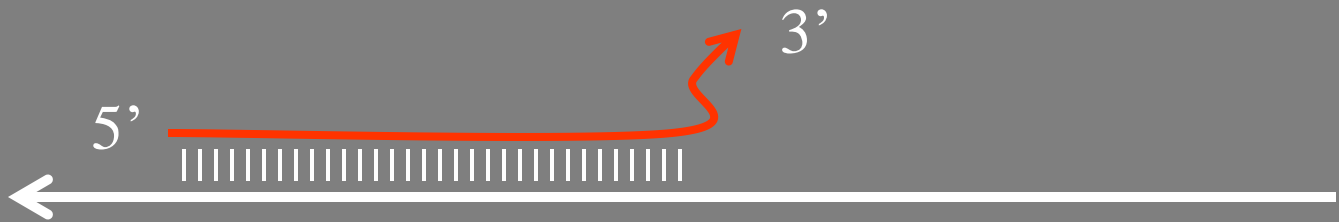
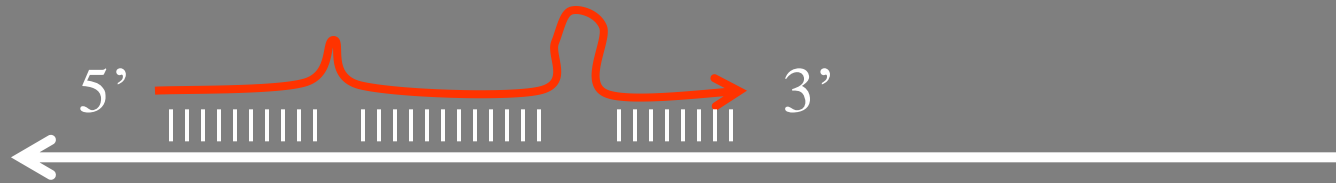
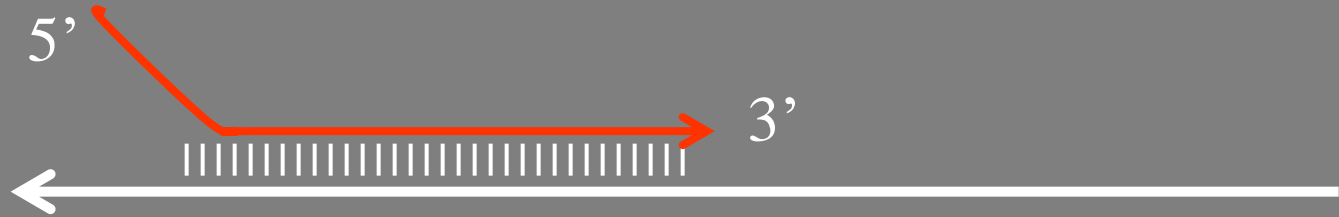
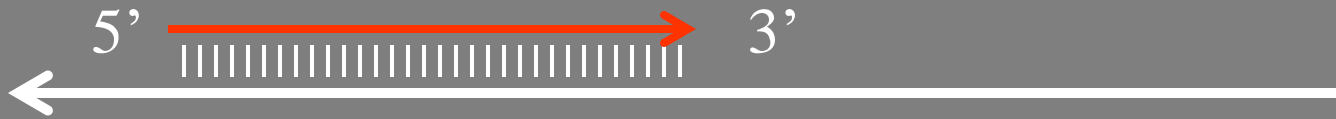


Specificity?

Proper annealing to the template?

- ▶ Mismatches towards the 3' end affect target amplification much more than mismatches towards the 5' end
 - ▶ a two base mismatch at the 3' end generally prevents amplification
 - ▶ A single base mismatch (even at the very 3' end), as well as a few mismatches in the middle or toward the 5' end, still allows amplification, though at a reduced efficiency for some cases
- 

When is a “primer” a primer?



Primer BLAST

Basic Local Alignment Search Tools

- ▶ Searching primers against an appropriate nucleotide sequence database
 - potential targets
 - number and the positions of matched bases
 - primer orientations and distance between forward and reverse primers
- ▶ time-consuming and very difficult task for users, especially when the primers have a large number of hits
- ▶ Similarity Searching using BLAST program has been widely used for primer target detection
 - to compare a query sequence against a sequence database

Now: Primer Blast

- ▶ to design target-specific primers for PCR
- ▶ **In-Silico PCR** and **Reverse ePCR** vs **Primer Blast**:
 - Do not design primers
 - Not sensitive enough
 - to detect targets with a significant number of mismatches (potentially amplifiable)

Primer-BLAST

- ▶ incorporates BLAST with a global alignment algorithm
- ▶ a full primer-target alignment
- ▶ sensitive enough to detect targets that have a significant number of mismatches to primers
- ▶ design new target-specific primers in one step
- ▶ check the specificity of pre-existing primers
- ▶ very sensitive in detecting potential amplification targets
- ▶ capability to place primers based on exon/intron boundaries
- ▶ and excluding SNP sites in primers

<http://www.ncbi.nlm.nih.gov/tools/primer-blast>

highly sensitive default parameters to find seq potentially amplifiable in PCR

- ▶ Detect a target with up to 35% mismatches to the primer sequence
- ▶ default BLAST expect value cutoff is 30,000 for the primer-only case (3000 times higher than the standard BLAST program default)
- ▶ the higher the expect value cutoff, the more sensitive the search
- ▶ word size of seven (standard BLAST uses 11),
- ▶ 50,000 for the maximum number of database sequences (standard BLAST uses 250)
- ▶ 1 for match reward to mismatch penalty ratio (standard BLAST uses 1.5)

Check primers specificity

- ▶ Go to the [Primer BLAST](#) submission form.
- ▶ Enter one or both primer sequences in the Primer Parameters section of the form
 - If only one primer is available, a template sequence is also required
- ▶ In the Primer Pair Specificity Checking Parameters section, select the appropriate source Organism and the smallest Database that is likely to contain the target sequence.
 - For broadest coverage, choose the nr database and do not specify an organism.
- ▶ Click the "Get Primers" button to submit the search and retrieve template and specificity information.

Primer BLAST submission form.

PCR Template

Reset page Save search parameters Retrieve recent results

Enter accession, gi, or FASTA sequence (A refseq record is preferred) Clear

Range

Forward primer From To Clear

Reverse primer Clear

Or, upload FASTA file Browse...

Primer Parameters

Use my own forward primer (5'→3' on plus strand) Clear

Use my own reverse primer (5'→3' on minus strand) Clear

PCR product size

of primers to return

Primer melting temperatures (T_m)

Min	Opt	Max	Max T _m difference
<input type="text"/>	<input type="text"/>	<input type="text"/>	<input type="text"/>

Exon/intron selection

A refseq mRNA sequence as PCR template input is required for options in the section

Exon junction span

Exon junction match

Exon at 5' side Exon at 3' side

Minimal number of bases that must anneal to exons at the 5' or 3' side of the junction

Intron inclusion Primer pair must be separated by at least one intron on the corresponding genomic DNA

Intron length range

Min	Max
<input type="text"/>	<input type="text"/>

Primer Pair Specificity Checking Parameters

Specificity check Enable search for primer pairs specific to the intended PCR template

Database

Organism

Enter an organism name, taxonomy id or select from the suggestion list as you type.

[Add more organisms](#)

Exclusion (optional) Exclude predicted Refseq transcripts (accession with XM, XR prefix) Exclude uncultured/environmental sample sequences

Entrez query (optional)

Primer specificity stringency

Primer must have at least total mismatches to unintended targets, including at least mismatches within the last bps at the 3' end.

Ignore targets that have or more mismatches to the primer.

Misprimed product size deviation

Splice variant handling Allow primer to amplify mRNA splice variants (requires refseq mRNA sequence as PCR template input)

Show results in a new window Use new graphic view

Enter one or both primer sequences in the Primer Parameters section of the form

Primer Pair Specificity Checking Parameters section

Selecting the appropriate source Organism and the smallest Database that is likely to contain the target sequence.

Click the "Get Primers" button to submit the search and retrieve template and specificity information

[Reset page](#) [Save search parameters](#) [Retrieve recent results](#)

PCR Template

Enter accession, gi, or FASTA sequence (A refseq record is preferred) [Clear](#)

Or, upload FASTA file

No file chosen

Range

	From	To	
Forward primer	<input type="text"/>	<input type="text"/>	Clear
Reverse primer	<input type="text"/>	<input type="text"/>	

Primer Parameters

Use my own forward primer (5'→3' on plus strand)
Use my own reverse primer (5'→3' on minus strand)

<input type="text" value="AGGTAGAAACGCCGGATCTCCTTACA"/>	Clear
<input type="text" value="GAGCTAACAGAGGATTTGGTAGGTGTGC"/>	Clear

PCR product size	Min <input type="text" value="70"/>	Max <input type="text" value="1000"/>
# of primers to return	<input type="text" value="5"/>	

Primer melting temperatures (T _m)	Min <input type="text" value="57.0"/>	Opt <input type="text" value="60.0"/>	Max <input type="text" value="63.0"/>	Max T _m difference <input type="text" value="3"/>
-----------------------------------------------	---------------------------------------	---------------------------------------	---------------------------------------	--------------------------------------------------------------

Exon/intron selection

A refseq mRNA sequence as PCR template input is required for options in the section

Exon junction span

No preference

Exon junction match

Exon at 5' side Exon at 3' side

7

4

Minimal number of bases that must anneal to exons at the 5' or 3' side of the junction

Intron inclusion

Primer pair must be separated by at least one intron on the corresponding genomic DNA

Intron length range

Min

1000

Max

1000000

Note: Parameter values that differ from the default are highlighted in yellow

Primer Pair Specificity Checking Parameters

Specificity check

Enable search for primer pairs specific to the intended PCR template

Database

Genome (reference assembly from selected organisms)

Organism

Homo sapiens

Enter an organism name, taxonomy id or select from the suggestion list as you type.

[Add more organisms](#)

Exclusion (optional)

Exclude predicted Refseq transcripts (accession with XM, XR prefix) Exclude uncultured/environmental sample sequences

Entrez query (optional)

Primer specificity stringency

Primer must have at least 2 total mismatches to unintended targets, including

at least 2 mismatches within the last 5 bps at the 3' end.

Ignore targets that have 6 or more mismatches to the primer.

Misprimed product size deviation

4000

Splice variant handling

Allow primer to amplify mRNA splice variants (requires refseq mRNA sequence as PCR template input)

Get Primers

Show results in a new window Use new graphic view

Advanced parameters

Note: Parameter values that differ from the default are highlighted in yellow

Checking the specificity of pre-existing primers

Primer pair 1

	Sequence (5'->3')	Length	Tm	GC%	Self complementarity	Self 3' complementarity
Forward primer	GTAGGACTGCTCAGTTCAAACAT	23	52.81	43.48	6.00	2.00
Reverse primer	ACAGTTACTACACCCGTAAGGC	22	53.90	50.00	5.00	2.00

Products on target templates

>[NM_001098494.1](#) Homo sapiens zinc finger protein 419 (ZNF419), transcript variant 5, mRNA

```
product length = 444
Forward primer 1 GTAGGACTGCTCAGTTCAAACAT 23
Template       493 ..... 515

Reverse primer 1 ACAGTTACTACACCCGTAAGGC 22
Template       936 ..... 915
```

```
product length = 780
Forward primer 1 GTAGGACTGCTCAGTTCAAACAT 23
Template       493 ..... 515

Reverse primer 1 ACAGTTACTACACCCGTAAGGC 22
Template       1272 .....CG..G...TT..... 1251
```

>[NM_001098491.1](#) Homo sapiens zinc finger protein 419 (ZNF419), transcript variant 1, mRNA

```
product length = 444
Forward primer 1 GTAGGACTGCTCAGTTCAAACAT 23
Template       592 ..... 614

Reverse primer 1 ACAGTTACTACACCCGTAAGGC 22
Template       1035 ..... 1014
```

```
product length = 780
Forward primer 1 GTAGGACTGCTCAGTTCAAACAT 23
Template       592 ..... 614

Reverse primer 1 ACAGTTACTACACCCGTAAGGC 22
Template       1371 .....CG..G...TT..... 1350
```

>[NM_198542.1](#) Homo sapiens zinc finger protein 773 (ZNF773), mRNA

```
product length = 444
Forward primer 1 GTAGGACTGCTCAGTTCAAACAT 23
Template       453 ....A.G...CA.....G. 475

Reverse primer 1 ACAGTTACTACACCCGTAAGGC 22
Template       896 .....C.....A..... 875
```

- ▶ Default specificity parameters
- ▶ Perfect matches to the ZNF419 gene transcript variant 5 and would generate a 444 base amplicon.
- ▶ Some other potential amplicons:
 - 780 base amplicon: in ZNF419 transcript
 - 444 base amplicon: a different gene (i.e., human zinc finger protein 773, Genbank accession NM_198542.1)
 - However, there are up to 5 mismatches between at least one of the primers and the targets, which is probably sufficient to prevent amplification interference or non-specific amplification
- ▶ Users can analyze this result and make judgment based on their own experimental experiences.

Comparison to other primer design tools

- ▶ offers the ability to specify the **number of mismatches** that a specific primer pair must have to unintended targets
- ▶ custom 3' end region: number of mismatches must be present
- ▶ the specificity of a primer is typically judged by the number of mismatches it has to unintended targets
 - higher number of mismatches offer more specificity
 - the locations of such mismatches (mismatches closer to 3' end offer more specificity)
- ▶ place primers on different exons (i.e., to span an intron) to avoid amplification of genomic DNA
- ▶ customization of the number of nucleotide matches on either side of an exon/exon junction
- ▶ Primer-BLAST presents detailed alignments between the primers and targets found

- ▶ high detection sensitivity
- ▶ capable of detecting potential amplification targets that have up to 5 mismatches to a primer
- ▶ by using highly sensitive BLAST parameters
- ▶ Primer-BLAST (with default parameters) will miss any targets that have 6 or fewer consecutive matches to a primer (since Primer-BLAST uses a word size of 7 by default).
 - For example, if a target has mismatches to a primer of 20 bases at positions 7 and 14 (assuming the 5' end is position one), the target will be missed by Primer-BLAST (with default parameters) even though it has only 2 mismatches.
 - Assuming a random distribution of mismatch locations, it is possible to calculate the number of possible arrangements of 18 matches and 2 mismatches.
 - There are 20×19 different ways to place the 2 mismatches among the 18 matches, but only 2 of these result in a word size shorter than 7, so the probability of missing a target with 2 mismatches to a primer of 20 bases is $2/(20 \times 19)$ or about 0.5%.

NCBI/ Primer-BLAST : results: Job id=JSID_01_88396_130.14.22.21_9003 [more...](#)

Input PCR template none
Specificity of primers Target templates were found in selected database: Genome database (reference assembly only) for selected species (Organism limited to Homo sapiens)
Other reports [▶ Search Summary](#)

▼ Detailed primer reports

Primer pair 1

	Sequence (5'→3')	Length	Tm	GC%	Self complementarity	Self 3' complementarity
Forward primer	AGGTAGAAACGCCGGATCTCCTTACA	26	65.17	50.00	7.00	1.00
Reverse primer	GAGCTAACAGAGGATTTGGTAGGTGTGC	28	65.25	50.00	5.00	2.00

Products on target templates

>[NT_030059.13](#) Homo sapiens chromosome 10 genomic contig, GRCh37.p10 Primary Assembly

product length = 241

Features associated with this product:

[cytochrome P450 2C9 precursor](#)

Forward primer	1	AGGTAGAAACGCCGGATCTCCTTACA	26
Template	47506464	...G.....C..	47506439
Reverse primer	1	GAGCTAACAGAGGATTTGGTAGGTGTGC	28
Template	47506224	47506251

Primer pair 1

	Sequence (5'>3')	Length	Tm	GC%	Self complementarity	Self 3' complementarity
Forward primer	AGGTAGAAACGCCGGATCTCCTTACA	26	65.17	50.00	7.00	1.00
Reverse primer	GAGCTAACAGAGGATTTGGTAGGTGTGC	28	65.25	50.00	5.00	2.00

Products on target templates

>[NT_030059.13](#) Homo sapiens chromosome 10 genomic contig, GRCh37.p10 Primary Assembly

product length = 241

Features associated with this product:

[cytochrome P450 2C9 precursor](#)

Forward primer	1	AGGTAGAAACGCCGGATCTCCTTACA	26
Template	47506464	...G.....C..	47506439
Reverse primer	1	GAGCTAACAGAGGATTTGGTAGGTGTGC	28
Template	47506224	47506251

product length = 239

Features associated with this product:

[cytochrome P450 2C19 precursor](#)

Forward primer	1	AGGTAGAAACGCCGGATCTCCTTACA	26
Template	47339662	...G.....C..	47339637
Reverse primer	1	GAGCTAACAGAGGATTTGGTAGGTGTGC	28
Template	47339424	47339451

► NCBI/ Primer-BLAST : results: Job id=JSID_01_88405_130.14.22.21_9003 [more...](#)

Input PCR template none
Specificity of primers Target templates were found in selected database: Genome database (reference assembly only) for selected species (Organism limited to Homo sapiens)
Other reports [► Search Summary](#)

▼ Detailed primer reports

Primer pair 1

	Sequence (5'→3')	Length	Tm	GC%	Self complementarity	Self 3' complementarity
Forward primer	AGGTAGAAACGCCGGATCTCCTTACA	26	65.17	50.00	7.00	1.00
Reverse primer	GTGTGTGCCTCTTTGATGGATAAA	24	59.54	41.67	4.00	4.00

Products on target templates

>[NT_022778.16](#) Homo sapiens chromosome 4 genomic contig, GRCh37.p10 Primary Assembly

product length = 733

Features flanking this product:

[64708 bp at 5' side: trans-2,3-enoyl-CoA reductase-like](#)

[849323 bp at 3' side: ephrin type-A receptor 5 isoform b precursor](#)

Reverse primer	1	GTGTGTGCCTCTTTGATGGATAAA	24
Template	5551175	..A.A.....T..A..A.....	5551152
Reverse primer	1	GTGTGTGCCTCTTTGATGGATAAA	24
Template	5550443	..A.A.....T..A..A.....	5550466

Input PCR template none
Specificity of primers Target templates were found in selected database: Genome database (reference assembly only) for selected species (Organism limited to Homo sapiens)
Other reports [► Search Summary](#)

▼ **Detailed primer reports**

Primer pair 1

	Sequence (5'→3')	Length	Tm	GC%	Self complementarity	Self 3' complementarity
Forward primer	AGAAGGAGCATATAGTGGGCCTAGGT	26	64.38	50.00	6.00	4.00
Reverse primer	GTGTGTGCCTCTTTGATGGATAAA	24	59.54	41.67	4.00	4.00

Products on target templates

>[NT_030059.13](#) Homo sapiens chromosome 10 genomic contig, GRCh37.p10 Primary Assembly

product length = 60

Features flanking this product:

[27127 bp at 5' side: cytochrome P450 2C18 isoform 2 precursor](#)

[76 bp at 3' side: cytochrome P450 2C19 precursor](#)

Forward primer	1	AGAAGGAGCATATAGTGGGCCTAGGT	26
Template	47326791	47326816
Reverse primer	1	GTGTGTGCCTCTTTGATGGATAAA	24
Template	47326850	47326827

product length = 60

Features flanking this product:

85636 bp at 5' side: cytochrome P450 2C19 precursor

74 bp at 3' side: cytochrome P450 2C9 precursor

Forward primer 1 AGAAGGAGCATATAGTGGCCTAGGT 26
Template 47502770A..... 47502795

Reverse primer 1 GTGTGTGCCTCTTTGATGGATAAA 24
Template 47502829 47502806

>[NT_022778.16](#) Homo sapiens chromosome 4 genomic contig, GRCh37.p10 Primary Assembly

product length = 733

Features flanking this product:

64708 bp at 5' side: trans-2,3-enoyl-CoA reductase-like

849323 bp at 3' side: ephrin type-A receptor 5 isoform b precursor

Reverse primer 1 GTGTGTGCCTCTTTGATGGATAAA 24
Template 5551175 ..A.A.....T..A..A..... 5551152

Reverse primer 1 GTGTGTGCCTCTTTGATGGATAAA 24
Template 5550443 ..A.A.....T..A..A..... 5550466

Primer pair 1

	Sequence (5'→3')	Length	Tm	GC%	Self complementarity	Self 3' complementarity
Forward primer	AGAAGGAGCATATAGTGGGCCTAGGT	26	64.38	50.00	6.00	4.00
Reverse primer	TATCTGTAGGATATTTCCAATCACTGGGA	29	61.32	37.93	6.00	5.00

Products on target templates

>[NT_030059.13](#) Homo sapiens chromosome 10 genomic contig, GRCh37.p10 Primary Assembly

product length = 271

Features associated with this product:

[cytochrome P450 2C19 precursor](#)

Forward primer	1	AGAAGGAGCATATAGTGGGCCTAGGT	26
Template	47326791	47326816
Reverse primer	1	TATCTGTAGGATATTTCCAATCACTGGGA	29
Template	47327061	47327033

product length = 269

Features associated with this product:

[cytochrome P450 2C9 precursor](#)

Forward primer	1	AGAAGGAGCATATAGTGGGCCTAGGT	26
Template	47502770A.....	47502795
Reverse primer	1	TATCTGTAGGATATTTCCAATCACTGGGA	29
Template	47503038	47503010

Primer pair 1

	Sequence (5'→3')	Length	Tm	GC%	Self complementarity	Self 3' complementarity
Forward primer	TTTCAAATGGGAAAAGGGAGACCCTGG	27	65.12	48.15	6.00	3.00
Reverse primer	TTCTCATGAGCATCTCTGGGGCTGTTTTCC	30	68.26	50.00	8.00	0.00

Products on target templates

>[NT_030059.13](#) Homo sapiens chromosome 10 genomic contig, GRCh37.p10 Primary Assembly

product length = 329

Features flanking this product:

[26264 bp at 5' side: cytochrome P450 2C18 isoform 2 precursor](#)

[670 bp at 3' side: cytochrome P450 2C19 precursor](#)

Forward primer	1	TTTCAAATGGGAAAAGGGAGACCCTGG	27
Template	47326256	47326230
Reverse primer	1	TTCTCATGAGCATCTCTGGGGCTGTTTTCC	30
Template	47325928	47325957

product length = 290

Features flanking this product:

[84809 bp at 5' side: cytochrome P450 2C19 precursor](#)

[671 bp at 3' side: cytochrome P450 2C9 precursor](#)

Forward primer	1	TTTCAAATGGGAAAAGGGAGACCCTGG	27
Template	47502232	47502206
Reverse primer	1	TTCTCATGAGCATCTCTGGGGCTGTTTTCC	30
Template	47501943T.....C..	47501972

more info

Primer-BLAST

Primer-Blast results

NCBI/ Primer-BLAST : results: Job id=JSID_01_88610_130.14.18.128_9003 [more...](#)

 Please correct the following errors: Specified right primer exceeds built-in maximum of 36.

	Sequence (5'→3')	Length	Tm	GC%	Self complementarity	Self 3' complementarity
Forward primer	TTTCAAATGGGAAAAGGGAGACCCTGG	27	65.12	48.15	6.00	3.00
Reverse primer	TTTCATGAGCATCTCTGGG	20	56.70	50.00	8.00	4.00

Products on target templates

>[NT_030059.13](#) Homo sapiens chromosome 10 genomic contig, GRCh37.p10 Primary Assembly

product length = 329

Features flanking this product:

[26264 bp at 5' side: cytochrome P450 2C18 isoform 2 precursor](#)

[670 bp at 3' side: cytochrome P450 2C19 precursor](#)

```
Forward primer 1          TTTCAAATGGGAAAAGGGAGACCCTGG 27
Template       47326256  ..... 47326230
```

```
Reverse primer 1          TTTCATGAGCATCTCTGGG 20
Template       47325928  ..... 47325947
```

product length = 290

Features flanking this product:

[84809 bp at 5' side: cytochrome P450 2C19 precursor](#)

[671 bp at 3' side: cytochrome P450 2C9 precursor](#)

```
Forward primer 1          TTTCAAATGGGAAAAGGGAGACCCTGG 27
Template       47502232  ..... 47502206
```

```
Reverse primer 1          TTTCATGAGCATCTCTGGG 20
Template       47501943  .....T 47501962
```


product length = 295

Features flanking this product:

20855 bp at 5' side: cytochrome P450 2C19 precursor

64620 bp at 3' side: cytochrome P450 2C9 precursor

Forward primer 1 TTTCAAATGGGAAAAGGGAGACCCTGG 27
Template 47437989T..G.T.... 47438015

Reverse primer 1 TTTCATGAGCATCTCTGGG 20
Template 47438283T.....T 47438264

>[NT_011512.11](#) Homo sapiens chromosome 21 genomic contig, GRCh37.p10 Primary Assembly

product length = 4870

Features flanking this product:

195895 bp at 5' side: protein C-ets-2 isoform 1

151939 bp at 3' side: proteasome assembly chaperone 1 isoform a

Reverse primer 1 TTTCATGAGCATCTCTGGG 20
Template 26057447 C.....G..T...G.... 26057428

Reverse primer 1 TTTCATGAGCATCTCTGGG 20
Template 26052578 .G...C.A..... 26052597

>[NT_008183.19](#) Homo sapiens chromosome 8 genomic contig, GRCh37.p10 Primary Assembly

product length = 1041

Features flanking this product:

17127 bp at 5' side: hydroxyacid-oxoacid transhydrogenase, mitochondrial

7130 bp at 3' side: uncharacterized protein C8orf46

Reverse primer 1 TTTCATGAGCATCTCTGGG 20
Template 47438283T.....T 47438264

>[NT_011512.11](#) Homo sapiens chromosome 21 genomic contig, GRCh37.p10 Primary Assembly

product length = 4870

Features flanking this product:

[195895 bp at 5' side: protein C-ets-2 isoform 1](#)

[151939 bp at 3' side: proteasome assembly chaperone 1 isoform a](#)

Reverse primer 1 TTTCATGAGCATCTCTGGG 20
Template 26057447 C.....G..T...G.... 26057428

Reverse primer 1 TTTCATGAGCATCTCTGGG 20
Template 26052578 .G...C.A..... 26052597

>[NT_008183.19](#) Homo sapiens chromosome 8 genomic contig, GRCh37.p10 Primary Assembly

product length = 1041

Features flanking this product:

[17127 bp at 5' side: hydroxyacid-oxoacid transhydrogenase, mitochondrial](#)

[7130 bp at 3' side: uncharacterized protein C8orf46](#)

Reverse primer 1 TTTCATGAGCATCTCTGGG 20
Template 19263154 GCAG...C..... 19263135

Reverse primer 1 TTTCATGAGCATCTCTGGG 20
Template 19262114 AA.A.....C....T 19262133

NCBI/ Primer-BLAST : results: Job id=JSID_01_90747_130.14.18.128_9003 [more...](#)

Input PCR template none
Specificity of primers Target templates were found in selected database: Genome database (reference assembly only) for selected species (Organism limited to Homo sapiens)
Other reports [▶ Search Summary](#)

▼ Detailed primer reports

Primer pair 1

	Sequence (5'→3')	Length	Tm	GC%	Self complementarity	Self 3' complementarity
Forward primer	AAATGAGGAACAATCAGGAGACGGCA	26	64.61	46.15	3.00	0.00
Reverse primer	ATCTTGGGATGCTGAGTGCCTGGAGT	26	67.61	53.85	3.00	1.00

Products on target templates

>[NT_004487.19](#) Homo sapiens chromosome 1 genomic contig, GRCh37.p10 Primary Assembly

product length = 793

Features associated with this product:

[filaggrin](#)

Forward primer	1	AAATGAGGAACAATCAGGAGACGGCA	26
Template	3774288	3774263
Reverse primer	1	ATCTTGGGATGCTGAGTGCCTGGAGT	26
Template	3773496	3773521

product length = 4684

Features associated with this product:

filaggrin

Forward primer	1	AAATGAGGAACAATCAGGAGACGGCA	26
Template	3774288	3774263
Reverse primer	1	ATCTTGGGATGCTGAGTGCCTGGAGT	26
Template	3769605	C.....C	3769630

product length = 1765

Features associated with this product:

filaggrin

Forward primer	1	AAATGAGGAACAATCAGGAGACGGCA	26
Template	3774288	3774263
Reverse primer	1	ATCTTGGGATGCTGAGTGCCTGGAGT	26
Template	3772524	C.....C.....C	3772549

product length = 3709

Features associated with this product:

filaggrin

Forward primer	1	AAATGAGGAACAATCAGGAGACGGCA	26
Template	3774288	3774263
Reverse primer	1	ATCTTGGGATGCTGAGTGCCTGGAGT	26
Template	3770580	C.....C.....C	3770605

product length = 1765

Features associated with this product:

filaggrin

Forward primer	1	AAATGAGGAACAATCAGGAGACGGCA	26
Template	3771369T	3771344
Reverse primer	1	ATCTTGGGATGCTGAGTGCCTGGAGT	26
Template	3769605	C.....C	3769630

product length = 3709

Features associated with this product:

filaggrin

Forward primer	1	AAATGAGGAACAATCAGGAGACGGCA	26
Template	3771369T	3771344
Reverse primer	1	ATCTTGGGATGCTGAGTGCCTGGAGT	26
Template	3767661	G.....C	3767686

product length = 790

Features associated with this product:

filaggrin

Forward primer	1	AAATGAGGAACAATCAGGAGACGGCA	26
Template	3771369T	3771344
Reverse primer	1	ATCTTGGGATGCTGAGTGCCTGGAGT	26
Template	3770580	C.....C.....C	3770605

product length = 4681

Features associated with this product:

filaggrin

Forward primer	1	AAATGAGGAACAATCAGGAGACGGCA	26
Template	3771369T	3771344
Reverse primer	1	ATCTTGGGATGCTGAGTGCCTGGAGT	26
Template	3766689	G.A.....C	3766714

product length = 2737

Features associated with this product:

filaggrin

Forward primer	1	AAATGAGGAACAATCAGGAGACGGCA	26
Template	3771369T	3771344
Reverse primer	1	ATCTTGGGATGCTGAGTGCCTGGAGT	26
Template	3768633	C.....C.T.....C	3768658

product length = 790

Features associated with this product:

filaggrin

Forward primer	1	AAATGAGGAACAATCAGGAGACGGCA	26
Template	3767478	T.....T	3767453
Reverse primer	1	ATCTTGGGATGCTGAGTGCCTGGAGT	26
Template	3766689	G.A.....C	3766714

product length = 1762

Features associated with this product:

filaggrin

Forward primer	1	AAATGAGGAACAATCAGGAGACGGCA	26
Template	3767478	T.....T	3767453
Reverse primer	1	ATCTTGGGATGCTGAGTGCCTGGAGT	26
Template	3765717	C.....C.....C	3765742

product length = 2734

Features associated with this product:

filaggrin

Forward primer	1	AAATGAGGAACAATCAGGAGACGGCA	26
Template	3767478	T.....T	3767453
Reverse primer	1	ATCTTGGGATGCTGAGTGCCTGGAGT	26
Template	3764745	C.....C.....C	3764770

product length = 3709

Features associated with this product:

filaggrin

Forward primer	1	AAATGAGGAACAATCAGGAGACGGCA	26
Template	3773313	T.....A.....T	3773288
Reverse primer	1	ATCTTGGGATGCTGAGTGCCTGGAGT	26
Template	3769605	C.....C	3769630

product length = 793

Features associated with this product:

filaggrin

Forward primer	1	AAATGAGGAACAATCAGGAGACGGCA	26
Template	3770397	T.....A.....T	3770372
Reverse primer	1	ATCTTGGGATGCTGAGTGCCTGGAGT	26
Template	3769605	C.....C	3769630

product length = 2737

Features associated with this product:

filaggrin

Forward primer	1	AAATGAGGAACAATCAGGAGACGGCA	26
Template	3770397	T.....A.....T	3770372
Reverse primer	1	ATCTTGGGATGCTGAGTGCCTGGAGT	26
Template	3767661	G.....C	3767686

product length = 790

Features associated with this product:

filaggrin

Forward primer	1	AAATGAGGAACAATCAGGAGACGGCA	26
Template	3773313	T.....A.....T	3773288
Reverse primer	1	ATCTTGGGATGCTGAGTGCCTGGAGT	26
Template	3772524	C.....C.....C	3772549

UCSC In Silico PCR

Testing Primer Specificity

In-Silico PCR:

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UCSC In-Silico PCR

Genome: Assembly: Forward Primer: Reverse Primer:

Max Product Size: Min Perfect Match: Min Good Match: **Flip Reverse Primer:**

- ▶ Select genome
- ▶ Enter primers
- ▶ Minimum 15 bases
- ▶ Flip reverse primer?
- ▶ Submit

Configuration Options

Genome and Assembly - The sequence database to search.

Forward Primer - Must be at least 15 bases in length.

Reverse Primer - On the opposite strand from the forward primer. Minimum length of 15 bases.

Max Product Size - Maximum size of amplified region.

Min Perfect Match - Number of bases that match exactly on 3' end of primers. Minimum match size is 15.

Min Good Match - Number of bases on 3' end of primers where at least 2 out of 3 bases match.

Flip Reverse Primer - Invert the sequence order of the reverse primer and complement it.

(note: the tool does not handle ambiguous bases at this time—don't use Ns)

UCSC In-Silico PCR

Genome:	Assembly:	Forward Primer:	Reverse Primer:	
Mouse	Jul. 2007	TGCACCACCAaCTGCTT	GGATGCAGGGATGATG	submit
Max Product Size: 50000	Min Perfect Match: 18	Min Good Match: 18	Flip Reverse Primer: <input type="checkbox"/>	

About In-Silico PCR

In-Silico PCR searches a sequence database with a pair of PCR primers, using an indexing strategy for fast performance.

Configuration Options

Genome and Assembly - The sequence database to search.

Forward Primer - Must be at least 15 bases in length.

Reverse Primer - On the opposite strand from the forward primer. Minimum length of 15 bases.

Max Product Size - Maximum size of amplified region.

Min Perfect Match - Number of bases that match exactly on 3' end of primers. Minimum match size is 15.

Min Good Match - Number of bases on 3' end of primers where at least 2 out of 3 bases match.

Flip Reverse Primer - Invert the sequence order of the reverse primer and complement it.

Output

When successful, the search returns a sequence output file in fasta format containing all sequence in the database that lie between and include the primer pair. The fasta header describes the region in the database and the primers. The fasta body is capitalized in areas where the primer sequence matches the database sequence and in lower-case elsewhere. Here is an example:

```
>chr22:31000551+31001000 TAACAGATTGATGATGCATGAAATGGG CCCATGAGTGGCTCCTAAAGCAGCTGC
TtACAGATTGATGATGCATGAAATGGGgggtggccaggggtgggggtga
gactgcagagaaaggcagggctggttcataaacaagctttgtgcgtcccaa
tatgacagctgaagttttccagggctgatggtgagccagtgagggttaag
tacacagaacatcctagagaaccctcattccttaagattaaaaataaa
```

About In-Silico PCR

- ▶ In-Silico PCR searches a sequence database with a pair of PCR primers, using an indexing strategy for fast performance.
- ▶ **Configuration Options**
 - **Genome and Assembly:** The sequence database to search
 - **Forward Primer:** Must be at least 15 bases in length
 - **Reverse Primer:** On the opposite strand from the forward primer. Minimum length of 15 bases.
 - **Max Product Size**
 - **Min Perfect Match:** Number of bases that match exactly on 3' end of primers. Minimum match size is 15.
 - **Min Good Match:** Number of bases on 3' end of primers where at least 2 out of 3 bases match.
 - **Flip Reverse Primer** - Invert the sequence order of the reverse primer and complement it.

- ▶ When successful, the search returns a sequence output file in **fasta format** containing all sequence in the database that lie between and include the primer pair.
- ▶ The **fasta header** describes the region in the database and the primers.
- ▶ The **fasta body** is capitalized in areas where the primer sequence matches the database sequence and in lower-case elsewhere.
- ▶ The + between the coordinates in the fasta header indicates this is on the positive strand.

In Silico PCR Results

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UCSC In-Silico PCR

Forward primer

Reverse primer

```
>chr22:32629059+32629508 450bp TAACAGATTGATGATGCATGAAATGGG CCCATGAGTGGCTCCTAAAGCAGCTGC  
TtACAGATTGATGATGCATGAAATGGGgggtggccaggggtgggggggtga  
gactgcagagaaaaggcagggctggtccataacaagctttgtgcgtcccaa  
tatgacagctgaagttttccaggggctgatggtgagccagtgaggtaag  
tacacagaacatcctagagaaaacctcattccttaagattaaaaataaa  
gacttgctgtctgt aagggattggattatcctatttgagaaattctgta  
tccagaatggcttaccacacaatgctgaaaagtgtgtaccgtaatctcaa  
agcaagctcctcctcagacagagaaaaccagcctgtcacaggaagcaaaag  
aaattggcttcacttttaaggtgaatccagaaccagatgtcagagctcc  
aagcactttgctctcagctccacGCAGCTGCTTTAGGAGCCACTCATGac
```

Match in uppercase

Mismatch in lowercase

Primer Melting Temperatures

Forward: 66.7 C taacagattgatgatgcatgaaatggg

Reverse: 73.8 C cccatgagtggctcctaaagcagctgc

The temperature calculations are done assuming 50 mM salt and 50 nM annealing oligo concentration. The code to calculate the melting temp comes from [Primer3](#).

Melting temperature



UCSC In-Silico PCR

No matches to aggtagaaacgccggatctccttaca aggtagaaacgccggatctccttaca in Human Feb. 2009 (GRCh37/hg19)

Primer Melting Temperatures

Forward: 67.8 C aggtagaaacgccggatctccttaca

Reverse: 67.8 C aggtagaaacgccggatctccttaca

The temperature calculations are done assuming 50 mM salt and 50 nM annealing oligo concentration. The code to calculate the melting temp comes from [Primer3](#).

UCSC In-Silico PCR

```
>chr10:96522327+96522386 60bp AGAAGGAGCATATAGTGGGCCTAGGT GTGTGTGCCTCTTTGATGGATAAA  
AGAAGGAGCATATAGTGGGCCTAGGTgattggccacTTTATCCATCAAAG  
AGGCACACAC
```

UCSC In-Silico PCR

```
>chr10:96522327+96522597 271bp AGAAGGAGCATATAGTGGGCCTAGGT TATCTGTAGGATATTTCCAATCACTGGGA  
AGAAGGAGCATATAGTGGGCCTAGGTgattggccactttatccatcaaag  
aggcacacacacttaattagcatggagtgttataaaaagcttggagtgca  
agctcacggttgtcttaacaagaggagaaggcttcaatggatccttttgt  
ggtccttggtgctctgtctctcatgtttgcttctcctttcaatctggagac  
agagctctgggagaggaaaactcctcctggccctactcctcTCCCAGTG  
ATTGGAAATATCCTACAGATA
```

UCSC In-Silico PCR

```
>chr10:96521464-96521792 329bp TTTCAAATGGGAAAAGGGAGACCCTGG TTCTCATGAGCATCTCTGGGGCTGTTTTCC  
TTTCAAATGGGAAAAGGGAGACCCTGGgagaacaggacacctgttggtgc  
cacacagctcatagctggcagaactgggatttgagctgaggtcttctgat  
gcccatcgtggcgcatatctcttacatcagagatgctttgagaacagaa  
gacacaaatttgaaaaaaaaaatcgtttgctaaaactttgttttagcaaa  
acaaaacaacttccaaacattagttattctgaatatataccacattcattc  
ctgttcataaaaacaggcttcacattaataagaaccacttatttatctaaG  
GAAAACAGCCCCAGAGATGCTCATGAGAA
```

Primer Melting Temperatures

Forward: 71.8 C tttcaaatgggaaaaggagaccctgg
Reverse: 74.4 C ttctcatgagcatctctggggctgttttcc

The temperature calculations are done assuming 50 mM salt and 50 nM annealing oligo concentration. The code to calculate the melting temp comes from [Primer3](#).

UCSC In-Silico PCR

No matches to agaaggaatatatagtgggcctaggt tatcaataggccatttccaatgactggg in Human Feb. 2009 (GRCh37/hg19)

Primer Melting Temperatures

Forward: 60.0 C agaaggaatatatagtgggcctaggt
Reverse: 69.0 C tatcaataggccatttccaatgactggg

The temperature calculations are done assuming 50 mM salt and 50 nM annealing oligo concentration. The code to calculate the melting temp comes from [Primer3](#).



UCSC In-Silico PCR

```
>chr10:96697479-96697768 290bp TTTCAAATGGGAAAAGGGAGACCCTGG TTTCATGAGCATCTCTGG
TTTCAAATGGGAAAAGGGAGACCCTGGgagaacaggacacctgttggtgc
cacacagctcatagctggcagaactgggatttgagctgaggtcttctgat
gcccatcgtggtgtattatctcttacaccagagctgccttgagaacaatt
ttagcaaaataaaacaaacttccaacattagttattctgaatatacacc
acatttattctgttcataaaaacaggcttcacattaaatagaacccttat
ttgtctctaagggaaacagcaCCAGAGATGCTCATGAGAA
>chr10:96521464-96521792 329bp TTTCAAATGGGAAAAGGGAGACCCTGG TTTCATGAGCATCTCTGG
TTTCAAATGGGAAAAGGGAGACCCTGGgagaacaggacacctgttggtgc
cacacagctcatagctggcagaactgggatttgagctgaggtcttctgat
gcccatcgtggcgcattatctcttacatcagagatgctttgagaacagaa
gacacaaatttgaaaaaaaaaatcgtttgctaaaactttgttttagcaaa
acaaaacaacttccaacattagttattctgaatataaccacattcatc
ctgttcataaaaacaggcttcacattaaatagaaccacttatttatctaag
gaaaacagcccCAGAGATGCTCATGAGAA
```

Primer Melting Temperatures

Forward: 71.8 C tttcaaatgggaaaaggagaccctgg

Reverse: 54.7 C ttctcatgagcatctctgg

The temperature calculations are done assuming 50 mM salt and 50 nM annealing oligo concentration. The code to calculate the melting temp comes from [Primer3](#).

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UCSC In-Silico PCR

```
>chr10:96697464-96697768 305bp TTCAAATGGGAAAAGGGAGACCCTGG GAGAACTCTTATTTTTTCTCATGAGCATCTCTGG
TTCAAATGGGAAAAGGGAGACCCTGGgagaacaggacacctggtggtgc
cacacagctcatagctggcagaactgggatttgagctgaggtcttctgat
gcccatcgtggtgtattatctcttacaccagagctgccttgagaacaatt
ttagcaaaataaaacaaacttccaaacattagttattctgaatatacacc
acatatttctgttcataaaaacaggcttcacattaatagaacccttat
ttgtctctaagggaaacagcaCCAGAGATGCTCATGAGAAAgAAATAAGgG
aTCTC
>chr10:96521449-96521792 344bp TTCAAATGGGAAAAGGGAGACCCTGG GAGAACTCTTATTTTTTCTCATGAGCATCTCTGG
TTCAAATGGGAAAAGGGAGACCCTGGgagaacaggacacctggtggtgc
cacacagctcatagctggcagaactgggatttgagctgaggtcttctgat
gcccatcgtggcgcatatctcttacatcagagatgctttgagaacagaa
gacacaaatttgaaaaaaaaaatcgtttgctaaaactttgtttagcaaa
acaaaacaacttccaaacattagttattctgaatataaccacattcattc
ctgttcataaaaacaggcttcacattaatagaaccacttatttcttaag
gaaaacagccCCAGAGATGCTCATGAGAAAAAATAAGAGTTCTC
```

Primer Melting Temperatures

Forward: 71.8 C tttcaaatgggaaaaggagaccctgg

Reverse: 68.4 C gagaactcttatttttctcatgagcatctctgg

The temperature calculations are done assuming 50 mM salt and 50 nM annealing oligo concentration. The code to calculate the melting temp comes from [Primer3](#).

Primer pair 1

	Sequence (5'→3')	Length	Tm	GC%	Self complementarity	Self 3' complementarity
Forward primer	TTTCAAATGGGAAAAGGGAGACCCTGG	27	65.12	48.15	6.00	3.00
Reverse primer	GAGAACTCTTATTTTTTCTCATGAGCATCTCTGG	34	64.09	38.24	8.00	2.00

Products on target templates

>[NT_030059.13](#) Homo sapiens chromosome 10 genomic contig, GRCh37.p10 Primary Assembly

product length = 344

Features flanking this product:

26249 bp at 5' side: cytochrome P450 2C18 isoform 2 precursor

670 bp at 3' side: cytochrome P450 2C19 precursor

Forward primer	1	TTTCAAATGGGAAAAGGGAGACCCTGG	27
Template	47326256	47326230
Reverse primer	1	GAGAACTCTTATTTTTTCTCATGAGCATCTCTGG	34
Template	47325913	47325946

product length = 305

Features flanking this product:

84794 bp at 5' side: cytochrome P450 2C19 precursor

671 bp at 3' side: cytochrome P450 2C9 precursor

Forward primer	1	TTTCAAATGGGAAAAGGGAGACCCTGG	27
Template	47502232	47502206
Reverse primer	1	GAGAACTCTTATTTTTTCTCATGAGCATCTCTGG	34
Template	47501928T.C.....C.....	47501961

	Sequence (5'→3')	Length	Tm	GC%	Self complementarity	Self 3' complementarity
Forward primer	TTTCAAATGGGAAAAGGGAGACCCTGG	27	65.12	48.15	6.00	3.00
Reverse primer	GAGAACTCTTATTTTTTCTCATGAGCATCTCTGGG	35	65.41	40.00	8.00	4.00

Products on target templates

>[NT_030059.13](#) Homo sapiens chromosome 10 genomic contig, GRCh37.p10 Primary Assembly

product length = 344

Features flanking this product:

[26249 bp at 5' side: cytochrome P450 2C18 isoform 2 precursor](#)

[670 bp at 3' side: cytochrome P450 2C19 precursor](#)

Forward primer	1	TTTCAAATGGGAAAAGGGAGACCCTGG	27
Template	47326256	47326230
Reverse primer	1	GAGAACTCTTATTTTTTCTCATGAGCATCTCTGGG	35
Template	47325913	47325947

product length = 305

Features flanking this product:

[84794 bp at 5' side: cytochrome P450 2C19 precursor](#)

[671 bp at 3' side: cytochrome P450 2C9 precursor](#)

Forward primer	1	TTTCAAATGGGAAAAGGGAGACCCTGG	27
Template	47502232	47502206
Reverse primer	1	GAGAACTCTTATTTTTTCTCATGAGCATCTCTGGG	35
Template	47501928T.C.....C.....T	47501962

UCSC In-Silico PCR

>[chr1:152281938-152285646](#) 3709bp AAATGAGGAACAATCAGGAGACGGCA ATCTTGGGATGCTGAGTGCCTGGAG

AAATGAGGAACAATCAGGAGACGGCAaccaggcactcagggtcacgtcacc
atgaagcttctctcaggetgacagctctagacactcacaggtgggcccag
ggacaatcaccggggcccaggacaagtaggaaccagggatccagtgttag
ccaggacagtgacagtcagggacactcagaagactctgagaggtggctctg
ggctctgcttccagaaaccatcatggatctgctcaggagcagtcagagat
ggctccagacaccccaggtcccacacgaagacagagctggctcatgggca
ctctgcagacagctccagaaaatcagggactcgtcacacacagaattcct
ctagtggacaggctgctgctcaccatgaacaggcaagatcaagtgcagga
gaaagacatggatcccggccaccagctccagtcagcagacagctccagaca
ctcaggcactgggacaggacaagcttcatctgcagtcagagacagtgagac
accgaggggtccagtggtagtcaggccactgacagtgagggacattcagaa
gactcagacacacagtcagtgctcaggccatggacaggctggtcaccatca
gcagagccaccaagagtcctgcacgtgaccggtcaggggaaaggtctcagc
gttcagggtcttctctaccagggtgagcactcataaacagctctgagtc
tcccatggatggacagggcccagcactggagtaagacaaggatcccacca
tgagcaggcacgagacaactccaggcactcagcatcccagatggtcagg
acaccattcgtggacacccgggggtcaagcagaagaggaaggcaggggtcc
caccacgagcaatcggtagataggtctggacactcaggggtcccacacag
ccacaccacatcccagggaaaggtctgatgcctcccgtgggcagtcaggat
ccagaagtgcaagcagaacaacacgtaaatgaggaacaatcaagagacggc
tccaggcactcagggtcacgtcaccatgaagcttctctcatgccgacat
ctctagacactcacaggcaggccagggacaatcagaggggtccaggacaa
gcaggcggccaggatccagtgtagccaggacagtgacagtgagggacat
tcagaagactctgagaggtggctctgggtctgcttccagaaaccatcgtgg



atagagccagtcattgggcactctgcagagagctccagacaatcaggcact
cgatcatgcagagacttctcttgggtggacaggctgcacatcccaggaaca
ggcaaggctcaagtcaccaggagaaagacatggatcccgccaccagcagtcag
cagacagctccacagactcaggcactggggcgcagacaagattcatctgta
gtcgggagacagtggaaccgagggtccagtggtagccaggccagtgacag
cgagggacactcagaagagtcagacacacagtcagtgtcagcccacggac
aggctgggccccatcagcagagccaccaagagtcacacagtgggccagtc
gggaaaggctctggacgttcagggtcttctcttaccaggtgagcactca
tgaacagtcctgagtcggccatggacgcacagggcccagcactggaggaa
gacaaagatcccgccacgagcaggcagcagagacagCTCCAGGCACTCAGCg
TCCCAAGAg

>chr1:152283882-152285646 1765bp AAATGAGGAACAATCAGGAGACGGCA ATCTTGGGATGCTGAGTGCCTGGAG

AAATGAGGAACAATCAGGAGACGGCAaccaggcactcagggtcacgtcatc
atgaagcttctctcaggttgacagctctagacactcacaggtggggccag
ggacaatcatcggggcccaggacaagtaggaaccagggatccagtgtag
ccaggacagtgacagtcagggacactcagaagactctgagaggtggtctg
ggtctgcttccagaaaccatcatggatctgctcaggagcagtcgaagat
ggctccagacaccccagggtccatcacgaagacagagctggtcattgggca
ctctgcagacagctccagaaaatcaggcactcgtcacacacagaattct
ctagtggacaggctgctgcatcccattgaacaggcaagatcaagtgcagga
gaaagacatggatcccgccaccagctccagtcagcagacagctccagaca
ctcaggcactggggcacggacaagcttcatctgcagtcagagacagtgagc
accgagggtccagtggttagtcaggccactgacagtgagggacattcagaa
gactcagacacacagtcagtgtcaggccatggacaggctggtcaccatca
gcagagccaccaagagtcggcacgtgaccggtcaggggaaaggctctgac
gttcagggtcttctcttaccaggtgagcactcataaacagctctgagtc
tccatggatggacaggggcccagcactggagtaagacaaggatcccacca
tgagcaggcacgagacaactccaggcactcagcatcccgaagatggtcagg
acaccattcgtggacacccgggggtcaagcagaagaggaaggcagggggtcc
caccacgagcaatcggtagataggctctggacactcagggtcccattcacag
ccacaccacatcccagggaaggctctgatgcctcccgtgggcagtcaggat
ccagaagtgaagcagaacaacacgtaattgaggaacaatcaagagacggc
tccagqactcagggtcacgtcaccatgaagcttctctctcatgccqacat

tctgcagtcagagacagtgacactgggggtccagtggttagtcaggccag
tgatagtgagggacattcagaggagtcagacacacagtcagtgtcagggc
atggacaggatgggccccatcagcagagccaccaagagtcctgcacgtgac
tggtcagggggaaggtctggacgttcaggggtctttcatctaccaggtgag
cactcatgaacagttctgagtcctgcccattggggcggaccaggaccagcactg
gacgaagacaaggatcccaccacgagcagggcacgagacagCTCCAGGCAC
TCAGCgTCCCAAGAg

>[chr1:152284854-152285646](#) 793bp AAATGAGGAACAATCAGGAGACGGCA ATCTTGGGATGCTGAGTGCCTGGAG

AAATGAGGAACAATCAGGAGACGGCAccaggcactcagggtcacgtcacc
atgaagcttcctctcagggtgacagctctagacactcacaggtgggcccag
ggacaatcatcggggcccaggacaagttaggaaccagggatccagtggttag
ccaggacagtgacagtcagggacactcagaagactctgagaggtggctctg
ggtctgcttccagaaaccatcatggatctgctcaggagcagtcgaagagat
ggctccagacaccccagggtcccattcacgaagacagagctggctcatgggca
ctctgcagacagctccagaaaatcaggcactcgtcacacacagaattcct
ctagtgagcaggctgctgcatcccattgaacaggcaagatcaagtgcagga
gaaagacatggatcccggccaccagctccagtcagcagacagctccagaca
ctcaggcactggggcaggacaagcttcattctgcagtcagagacagtgagc
accgaggggtccagtggttagtcaggccactgacagtgagggacattcagaa
gactcagacacacagtcagtgctcaggccattggacagggtgggtcaccatca
gcagagccaccaagagtcctgcacgtgaccgggtcaggggaaaggtctcgac
gttcaggggtctttcctctaccaggtgagcactcataaacagttctgagtc
tcccatggatggacagggcccagcactggagtaagacaaggatcccacca
tgagcagggcacgagacaaCTCCAGGCACTCAGCATCCCAAGAT

Primer Melting Temperatures

Forward: 69.7 C aaatgaggaacaatcaggagacggca

Reverse: 70.6 C atcttgggatgctgagtgcctggag

The temperature calculations are done assuming 50 mM salt and 50 nM annealing oligo concentration.

[Primer3](#).